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Genetic variance, coefficient of parentage, and genetic distance of six soybean populations

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Abstract Plant breeders would like to predict which biparental populations will have the largest genetic variance. If the population genetic variance could be predicted using coefficient of parentage or genetic distance estimates based on molecular marker data, breeders could choose parents that produced segregating populations with a large genetic variance. Three biparental soybean [*Glycine max* (L.) Merr.] populations were developed by crossing parents that were closely related, based on pedigree relationships. Three additional biparental populations were developed by crossing parents that were assumed to be unrelated. The genetic variance of each population was estimated for yield, lodging, physiological maturity, and plant height. Coefficient of parentage was calculated for each pair of parents used to develop the segregating populations. Genetic distance was determined, based on the number of random amplified polymorphic markers (RAPD) that were polymorphic for each pair of parents. Genetic distance was not associated with the coefficient of parentage or the magnitude of the genetic variance. The genetic variance pooled across the three closely related populations was smaller than the genetic variance pooled across the three populations derived from crossing unrelated parents for all four traits that were evaluated.

Key words *Glycine max* · DNA · Pedigree analysis · Genetic variance

Introduction

Plant breeders that develop segregating populations from biparental crosses would like to predict the mean and genetic variance of experimental lines derived from each two-parent combination. Populations with a high mean and large genetic variance would be expected to be the most useful for developing improved cultivars.

Prediction of the population mean from the mid-parent mean has been evaluated by several investigators. Busch et al. (1974) found that the mean yield of 25 populations of experimental lines of wheat (*Triticum aestivum* L.) was highly correlated ($r = 0.73$, $P = 0.01$) with mid-parent means. Souza and Sorrells (1991) found that means of three traits were not significantly different than the mid-parent means for 20 oat populations. In the oat populations studied by Cowan and Frey (1987), the mid-parent mean generally was higher than the progeny mean.

Predicting the genetic variance within a biparental population is more difficult and less reliable than predicting the population mean. The coefficient of parentage (Kempthorne 1969) has been suggested as one measure of genetic diversity. However, the coefficient of parentage requires simplifying assumptions and quantifies only the probability of two alleles being identical by descent. Breeders are perhaps more concerned with the probability of two alleles being alike in state, because the genetic variance among progeny would be determined by the number of alleles that are alike in state.

The association between the genetic variance and measures of genetic diversity among the parents has been evaluated. Souza and Soorrells (1991) determined that the coefficient of parentage was a better predictor of genetic variances than similarity indexes derived from discrete morphological or biochemical markers. Cowen and Fry (1987) found a significant correlation ($r = 0.41$) between the log of the generalized genetic variance and genealogical distance.

Cox et al. (1985a) suggested that the ideal breeding population should have a high mean for the traits of

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interest and a high genetic variance. They further suggested that a similarity index which combines coefficient of parentage and data derived from biochemical and morphological markers could be a useful measure of genetic diversity. In their analysis of soybean cultivars released after 1970, they found a correlation of $r = 0.60$ ($P = 0.1$) between coefficient of parentage and a similarity index measured with morphological and biochemical markers.

In a separate study Cox et al. (1985b) compared the coefficient of parentage of selected hard red winter wheat (*Triticum aestivum* L) parents to a similarity index derived from polyacrylamide gel electrophoresis data of gliadin storage protein patterns. Because the ancestors of hard red winter wheat were heterogeneous landraces, the correlation between these two parameters was only 0.27 ($P = 0.01$).

If the mean and genetic variance of biparental populations could be predicted, the best two parent combinations could be selected for crossing. Either the coefficient of parentage or biochemical markers might be used to predict the genetic similarity between parents. Our objectives were to: (1) compare the coefficient of parentage to the genetic distance estimated from random amplified polymorphic DNA (RAPD) data; (2) compare the genetic distance estimated from RAPD markers to the genetic component of variance estimate; (3) compare the coefficient of parentage to the genetic component of variance estimate for populations derived from closely and distantly related parents.

Materials and methods

Genetic relationships

Six soybean populations were developed, three from crosses among parents that were closely related as determined by published pedigree relationships and three additional populations among unrelated parents. The coefficient of parentage among these unrelated parents was assumed to be zero based on pedigree relationships.

Cultivars 'Evans' (Lambert and Kennedy 1975) and 'Wilkin' (Lambert and Kennedy 1973) are derived from the cross of 'Merit' (Johnson 1960) and 'Harosoy' (Weiss and Stevenson 1955). The cultivar 'Ozzie' (Orf et al. 1985) is derived from a cross of 'Wilkin' × M63-217Y. M63-217Y is a yellow hilum selection from 'Hodgson' (Lambert and Kennedy 1975). 'Pioneer 9061' has the pedigree 'Wells' × ('Corsoy' / 2 × 'Rampage') (Wilcox et al. 1973; Weber and Fehr 1970a; Weber and Fehr 1970b) and 'Pioneer 0877' has the pedigree 'Corsoy' × ('Clark' × 'Chippewa 64') (Johnson 1958; Bernard 1964). The experimental line ND867 is a North Dakota State University-derived line with the pedigree 'Wilkin' × L62-361. L62-361 is a Dt2 semi-determinate near-isogenic derivative of 'Harosoy'. 'Ozzie', 'Evans', 'Pioneer 9061', and 'Pioneer 0877' are Maturity Group 0 cultivars. 'Wilkin' and 'ND867' are Maturity Group 00 genotypes. These genotypes are considered to be adapted and high yielding.

PI 238924 is a Maturity Group 0 accession from Czechoslovakia with the name 'Kirches Stamm 2008' and was added to the USDA Soybean Germplasm Collection in 1957 (Bernard et al. 1989). PI417511 is a Maturity Group 000 accession from France with the name 'Grignon 48' and was added to the USDA Soybean Germplasm Collection in 1977 (Bernard et al. 1989). PI 261475 is a Maturity Group 0 accession from northeast China with the name 'Shika No. 1' and was added to the USDA Soybean Germplasm Collection in 1959 (Bernard et al. 1989). None of these plant introductions (PI) are in the

pedigrees of the adapted cultivars or experimental lines used as parents in this experiment.

The following crosses were made in the summer of 1989 at Fargo, ND: 'Ozzie' × 'Wilkin', 'Wilkin' × 'Evans', 'Ozzie' × 'ND867', 'Evans' × PI 417511, 'Pioneer 9061' × PI 238924, and 'Pioneer 0877' × PI 261475. The resulting F_1 plants were grown in the winter of 1989–1990 in Chile. The F_2 populations were grown in the 1990 Fargo summer nursery. The F_3 populations were grown in the 1990–1991 Chile winter nursery, and $F_{3.4}$ lines were developed by single-plant threshing approximately 100 individual plants from each population. No selection was applied to these populations, and the experimental lines derived from each population are random lines. The $F_{3.4}$ lines were evaluated in progeny rows in the summer of 1991. Ninety-nine lines were harvested from each population with the exception that 94 lines were derived from the population 'Pioneer 0877' × PI 261475. The experimental lines were evaluated for physiological maturity in 1991, based on data from a single progeny row.

Experimental design and statistical analysis

Eleven sets were formed by grouping lines of similar maturity into a set. Each set consisted of 9 lines from each population, except for the latest maturing set. The set of latest maturing lines contained 9 lines from five populations and 4 lines from the 'Pioneer 0877' × PI 261475 population. Each set was arranged in a square lattice design and included 54 experimental lines, the nine parental genotypes and a Maturity Group 00 cultivar 'McCall'. 'McCall' was used to complete the required 64 entries for an 8×8 lattice experiment.

Each set was arranged in a separate 8×8 square lattice design to form 11 lattice sets per environment. Each lattice design was replicated twice at each location. The experiment was evaluated at Casselton, N.D. and Morris, Minn. in 1993 and 1994. Plots at Casselton, N.D. had a planted length of 6.4 m. The row spacing within and between plots was 0.76 m and the center 4.6 m of a two-row plot was harvested. Plots at Morris, Minn. had a planted length of 3 m with a between-row spacing of 0.25 m. The center 2.4 m of a four-row plot was harvested.

Grain yield, plant height, physiological maturity, and lodging were evaluated. Grain yield was converted to kilograms per hectare at 13% water content. Plant height, measured at maturity, was recorded as the distance (in centimeters) from the base of the plant to the terminal node. Dates when 95% of the pods were the mature plant color were recorded at all locations, except Casselton in 1993. Grain yield and plant height were evaluated in all four environments. Lodging was evaluated at Casselton and Morris in 1994. In 1993, there was not an appreciable amount of lodging at either location.

Within each environment and set the data were adjusted for lattice block effects using the Statistical Analysis Package (SAP) computer program (Hammond 1993). The adjusted means of all environments were included in a combined analysis across environments, and entries were pooled across sets. Environments, lines and sets were considered to be random effects. Populations were considered to be fixed effects. Mean squares were equated to expected mean squares, and the genetic component of variance was calculated for each population. Standard errors of estimates for the genetic variance components were calculated, based on the formula provided by Hallauer and Miranda (1981).

Further statistical analyses included pooling the sum of squares from the three closely related populations into group one. The sum of squares from the three populations derived from the crosses of the adapted × PI parents were pooled into group two. The sums of squares for the genotype × environment interaction were pooled across the three populations within each group. Lines derived from each population were nested within that population. Three populations were nested within each group. The population mean was subtracted from the mean of each line within that population before squaring and summing these deviations. These sums of squares for each population within a group were then pooled and divided by the pooled degrees of freedom.

The mean square for populations within each group was then equated to the expected mean square, and the genetic component of variance was estimated. The standard error of the genetic variance for

each group was calculated, based on the formula provided by Hallauer and Miranda (1981). When the difference between group-one and group-two genetic variance components was greater than the sum of the standard errors, the genetic variance components were considered to be significantly different.

This experiment was designed to compare populations derived from closely related parents to populations derived from unrelated parents. Each group of three populations were considered to be two levels (groups) of a genetic similarity factor. The coefficients of parentage were derived using a computer program provided by Cox et al. (1986). Cox et al. (1985b) stated the assumptions used for the calculations of the coefficients of parentage.

RAPD procedures

DNA was isolated from leaf material of each genotype by the technique of Doyle and Doyle (1990). The DNA was amplified using Operon Technology random decamer synthetic oligonucleotide primers using a modification of the procedure described by Martin et al. (1991). The reactions contained 20 ng template DNA, 400 nM primer, 20 mM Tris-HCl, 50 mM KCl, 1.5 mM MgCl₂, 200 nM each dNTP, and 1.0 unit *Taq* DNA polymerase in a reaction volume of 10 µl. The thermal cycle used was: 44 cycles at 94°C (1 min) for DNA denaturation, 35°C (1 min) to anneal primers, and 72°C (2 min) for primer extension, followed by a final cycle of 94°C (1 min), 35°C (1 min), and 72°C (7 min) for final extension. After the final cycle, samples were held at 4°C until analysis. Fragments generated by amplification were separated according to size on 2% agarose gels run in 1 × TAE (40 mM Tris-Acetate, pH 9.0, 1.0 mM EDTA), stained with ethidium bromide and visualized by illumination with ultraviolet light. A total of 80 primers were sampled, which resulted in 470 markers, of which 277 were polymorphic. One genotype, 'Pioneer 9061', was included twice in each amplification set as an internal check to evaluate the repeatability of the banding patterns. The duplicate lanes of 'Pioneer 9061' showed complete agreement for all marker bands.

Genetic distance

A pairwise comparison of the RAPD pattern of the two parental genotypes for each population over all primers was made, and the

number of products shared by the two genotypes and the number of unique products for each genotype were counted. From this data the proportion of products that was shared by the two genotypes (S_{xy}) was determined to estimate the genetic similarity. The following formula developed by Nei and Li (1979) was used:

$$S_{xy} = 2n_{xy}/(n_x + n_y)$$

where n_x and n_y are the total number of amplification products obtained with the 80 primers for genotypes X and Y, respectively. n_{xy} is the number of amplification products shared between the two parents.

The genetic distance (D_{xy}) is the proportion of products that are not in common between two genotypes X and Y and was defined $1 - S_{xy}$. The standard error (SE) of D_{xy} was calculated as: (Keim et al. 1992)

$$SE(D_{xy}) = [S_{xy}(D_{xy})/n]^{1/2}$$

where n is the number of markers. Markers that were monomorphic across all nine genotypes were excluded from the calculation of genetic distance. There were 277 non-monomorphic markers. If the sum of the standard errors of two genetic distances was smaller than the difference between the D_{xy} estimates between two combinations of two parents, then the difference was considered significant. A bootstrap technique was used to determine the number of polymorphic markers necessary to evaluate genetic distance. There was a negligible change in the genetic distance when calculations using 80% of the marker data were compared to 100% of the marker data.

Results and discussion

Mean yield of the 'Evans*Wilkin' population was higher than that of any of the three adapted × PI populations (Table 1). However, the 'Pioneer 0877'*PI 261475 population was higher yielding than the 'Ozzie'*ND867 population. The results show that adapted × PI populations can be developed that yield as much as

Table 1 Mean yield, plant height, physiological maturity, and lodging of six bi-parental derived soybean populations and parents

Population or parent	Yield (kg/ha)	Plant height (cm)	Physiological maturity (days ^a)	Lodging score ^b
Ozzie	2690	65	47	1.1
Wilkin	2610	62	46	1.2
Evans	3130	74	51	1.9
ND867	2060	61	39	1.4
PI 417511	1370	48	40	1.5
Pioneer 0877	2790	69	54	2.5
PI 261475	2670	65	51	3.1
Pioneer 9061	3190	65	50	1.2
PI 238924	2110	56	48	2.3
Ozzie*Wilkin	2710	65	48	1.3
Evans*Wilkin	2820	68	49	1.5
Ozzie*ND867	2580	68	50	1.4
PI 417511*Evans	2460	65	52	2.1
Pioneer 0877*				
PI 261475	2650	68	54	2.6
Pioneer 9061*				
PI 238924	2590	60	50	1.7
LSD (0.05) ^c	353	6	3	0.7
LSD (0.05) ^d	36	1	0.3	0.1

^a Days from August 1

^b Lodging score, 1 (all plants upright) to 5 (all plants prostrate)

^c Least significant difference between parents at the 0.05 probability

level

^d Least significant difference between population means at the 0.05 probability level

adapted \times adapted population crosses. All six populations were adapted to this northern USA soybean growing region. The maturity among these populations only varied by a range of 6 days. Populations that are unadapted due to late maturity may not express their yield potential in northern environments.

There was no apparent relationship between the coefficient of parentage and the genetic distance, based on RAPD markers (Table 2). For example, 'Evans' and 'Wilkin' had a coefficient of parentage of 0.53, and the genetic distance between these two parents was 0.29. 'Evans' and PI 417511 have an assumed coefficient of parentage of zero, and the genetic distance between these two parents was 0.31. 'Ozzie' and 'Wilkin'; 'Ozzie' and ND867; and 'Pioneer 9061' and PI238924 all have a similar genetic distance. However, the coefficient of parentage of 'Pioneer 9061' and PI 238924 is very different from that of 'Ozzie' and 'Wilkin' or 'Ozzie' and ND867.

The results suggest that coefficient of parentage and genetic distance are different measures of genetic diversity between two genotypes. Coefficient of parentage is a

Table 2 Coefficient of parentage and genetic distance and standard error (SE) of genetic distance for six combinations of two parents. Genetic distance was determined by the proportion of markers that were not common to the two parents. Monomorphic bands for all nine genotypes were excluded from genetic distance calculations

Parental combination	Coefficient of parentage	Genetic distance ^a	SE
Ozzie, Wilkin	0.63	0.14	0.02
Evans, Wilkin	0.53	0.29	0.02
Ozzie, ND867	0.49	0.17	0.02
Evans, PI 417511	0	0.31	0.03
Pioneer 0877, PI 261475	0	0.23	0.02
Pioneer 9061, PI 238924	0	0.20	0.02

^a Based on 277 polymorphic RAPD markers from 80 primers

measure of identity by descent. Genetic distance measured with RAPD data is a measure of alike in state for predominantly non-coding regions of the genome. Because the soybean genome consists of approximately 40% single-copy and 60% repetitive DNA (Goldberg 1978), the DNA markers should be distributed in the same ratio among these two DNA classes. Although the single-copy DNA and repetitive DNA classes contain expressed sequences, the majority of the genome complexity is not associated with expressed genes (Goldberg et al. 1978). Therefore, the majority of RAPD markers would be associated with non-expressed and neutral sequences that should not be affected by selection. Thus, the marker data as a whole should be reflective of the genetic distance among two parents.

It is important to our conclusions that we can detect significant differences between genetic variances. Genetic components of variance were estimated by equating mean squares (Table 3) to expected mean squares and solving. We defined a difference between two genetic variance components to be significant when the difference was greater than the sum of the standard errors.

Genetic distance was not an indication of genetic variance. The genetic distance of 'Ozzie' and 'Wilkin' was 0.14 (Table 2) and was similar to the genetic distance of 'Ozzie' and ND867, which was 0.17. However, the genetic variance estimate for yield of the 'Ozzie' \times 'Wilkin' population was less than one-half the magnitude of the genetic variance estimate for the 'Ozzie' \times ND867 population (Table 4). Additionally, 'Evans' and 'Wilkin' had a similar genetic distance as 'Evans' and PI 417511 (Table 2). However, the genetic variance estimate for yield of the 'Evans' \times 'Wilkin' population is about one-sixth as large as the genetic variance estimate of the 'Evans' \times PI 417511 population (Table 4).

The results show that genetic distance measured with RAPD markers was not a good indication of genetic

Table 3 Mean squares for six populations (POP) and two groups for both genotype and genotype \times environment (G \times E) for four traits, pooled across sets. Each group includes three populations, and each population includes approximately 99 inbred soybean lines

Source	Mean square			
	Yield	Plant height	Physiological maturity	Lodging
Genotype				
Group 1 ^a	249 238	105.02	37.32	0.20
POP 1 ^b	217 354	82.02	29.47	0.07
POP 2	167 723	94.16	23.70	0.23
POP 3	360 785	138.63	58.97	0.31
Group 2	384 178	138.05	49.78	0.66
POP 4	390 416	138.23	59.54	0.72
POP 5	260 732	126.81	40.04	0.70
POP 6	495 639	148.59	49.43	0.56
G \times E				
Group 1	122 118	42.46	5.99	0.11
POP 1	119 301	21.91	4.74	0.07
POP 2	121 189	43.00	5.80	0.11
POP 3	125 934	63.13	7.46	0.14
Group 2	152 188	33.57	8.13	0.25
POP 4	130 261	33.33	7.73	0.26
POP 5	141 980	36.62	8.99	0.25
POP 6	183 246	30.93	7.72	0.25

^a Group 1 includes populations 1, 2, and 3; group 2 includes populations 4, 5, and 6

^b Population 1 is 'Ozzie' \times 'Wilkin', population 2 is 'Evans' \times 'Wilkin', population 3 is 'Ozzie' \times ND867, population 4 is 'Evans' \times PI 417511, population 5 is 'Pioneer 0877' \times PI 261475, population 6 is 'Pioneer 9061' \times PI 238924

Table 4 Genetic variance component estimates, standard errors (SE), and number of environments evaluated for yield, plant height, physiological maturity, and lodging for six soybean populations

Trait	Population	Genetic variance Component	SE	Number of environments
<i>Yield</i>	Ozzie*Wilkin	24 500	8 600	4
	Evans*Wilkin	11 600	6 900	4
	Ozzie*ND867	58 700	13 800	4
	Evans*PI 417511	65 000	15 000	4
	Pioneer 0877*			
	PI 261475	29 700	10 500	4
	Pioneer 9061*			
	PI 238924	78 100	19 100	4
<i>Plant height</i>	Ozzie*Wilkin	15.0	3.1	4
	Evans*Wilkin	12.8	3.7	4
	Ozzie*ND867	18.9	5.4	4
	Evans*PI 417511	26.2	5.2	4
	Pioneer 0877*			
	PI 261475	22.6	5.0	4
	Pioneer 9061*			
	PI 238924	29.4	5.6	4
<i>Physiological maturity</i>	Ozzie*Wilkin	8.2	1.5	3
	Evans*Wilkin	6.0	1.2	3
	Ozzie*ND867	17.2	3.0	3
	Evans*PI 417511	17.3	3.0	3
	Pioneer 0877*			
	PI 261475	10.4	2.1	3
	Pioneer 9061*			
	PI 238924	13.9	2.5	3
<i>Lodging</i>	Ozzie*Wilkin	0.00	0.01	2
	Evans*Wilkin	0.06	0.02	2
	Ozzie*ND867	0.08	0.03	2
	Evans*PI 417511	0.23	0.06	2
	Pioneer 0877*			
	PI 261475	0.22	0.06	2
	Pioneer 9061*			
	PI 238924	0.16	0.05	2

variance for yield among these six populations. Also, there is no relationship between genetic distance and genetic variance for plant height, physiological maturity, or lodging among these six populations.

If the coefficient of parentage was useful for predicting the magnitude of the genetic variance component, genetic variance components of each of the three populations derived from crossing closely related parents would be expected to be of similar magnitude. However, a comparison of the genetic variance estimates for yield among the three populations derived from crossing closely related parents showed that this was not the case (Table 4). There was five-fold increase in genetic variance for yield between the 'Evans' \times Wilkin population as compared to the 'Ozzie' \times ND867 population. Therefore, coefficient of parentage was not a good predictor of genetic variance for a specific population derived from the cross of two parents.

Genetic variance estimates for plant height within each of the three populations derived from crossing closely related parents are similar in magnitude (Table 4). However, for physiological maturity and lodging, there are significant differences in genetic variance estimates among the three populations derived

from crossing closely related parents. When we compare the three populations derived from crossing closely related parents, there is no relationship between the coefficient of parentage and the genetic variance estimate for yield, physiological maturity, and lodging.

The apparent lack of relationship between coefficient of parentage and genetic variance is limited to these populations, and a closer association could occur among another set of populations. Additional research that includes different populations will be required to determine the overall association between the coefficient of parentage and genetic variance estimates. Due to the amount of resources required to estimate genetic variance components with sufficient precision for comparing populations, this experiment was not intended to provide a global answer to this question. However, it is apparent that the coefficient of parentage was not useful as a predictor of the magnitude of the genetic variance for a specific population derived from crossing two parents.

The explanation for the lack of relationship between the coefficient of parentage and the genetic variance is that the coefficient of parentage is a measure of the degree to which two genotypes (parents) are identical by

descent. The genetic variance is a measure of the degree to which two genotypes (parents) are alike in state. The coefficient of parentage estimates the minimum proportion of alleles that would be expected to be identical by descent and therefore alike in state between two parents. Also, of the alleles that are not identical by descent, the proportion of alleles that are alike in state is unknown. Another explanation for the lack of relationship between coefficient of parentage and genetic variance could be inaccurate pedigree information. Although pedigree relationships may not always be accurately reported, the applied plant breeder must use the pedigree information that is available.

Among these six populations, the genetic variance component for yield in the 'Ozzie' × ND867 population was larger than the genetic variance component for yield of the 'Pioneer 0877' × PI 261475 population (Table 4). The genetic variance for plant height within the 'Pioneer 0877' × PI 261475 population was not significantly different than that of the 'Ozzie' × ND867 population. This result provides further evidence that the genetic variance within some populations derived from closely related parents are greater or equal to that of populations derived from unrelated parents. The genetic variance for physiological maturity of the 'Ozzie' × ND867 population is not significantly different from that of the 'Evans' × PI 417511 population, and the magnitudes of these estimates are very similar.

A comparison of the genetic variance for lodging among closely related and unrelated crosses provides a completely different conclusion than for the traits yield, plant height and physiological maturity (Table 4). The genetic variance for lodging of all three populations derived from closely related parents is smaller than that of any of the unrelated populations. The adapted × PI populations had a greater lodging susceptibility than the adapted × adapted populations (Table 1). The larger genetic variance component for lodging in the adapted × PI populations was probably due to the addition of lodging susceptible genes from the PI parents.

The genetic variance for yield, pooled across populations within each group, was significantly different between groups (Table 5). The pooled genetic variance for

populations developed from closely related crosses (group one) is smaller than that of populations developed from unrelated crosses (group two) for each of the four traits evaluated.

These results suggest that although breeders cannot predict the magnitude of the genetic variance component for any specific population from the coefficient of parentage, crosses among unrelated parents will, on average, result in a larger genetic variance than crosses among related parents. If the mean of two different populations comprised of inbred progenies are equal, the population with the larger genetic variance would be expected to produce a greater proportion of high yielding inbred lines. It will require further research, evaluating different populations, to determine whether this result is specific to this sample of populations.

Souza and Sorrells (1991) found that as the coefficient of parentage increased, the genetic variance estimate decreased for one trait of the six traits they evaluated. They found that crosses between more distantly related parents produced larger genetic variance estimates than crosses between closely related parents for plant biomass. However, crosses between closely related parents resulted in larger genetic variances than crosses among distantly related parents for grain yield, test weight, heading date, grain filling period, and maturity date. Because the crosses to the unadapted parents matured too late for complete expression of yield, the genetic variance estimates for yield were inconclusive.

The results of this experiment provide evidence that there was no relationship between the coefficient of parentage and genetic distance estimated from RAPD data. Also, there was no relationship between the genetic distance and the genetic component of variance estimate. The RAPD markers we used may not have been linked to any of the traits that were evaluated. In the future, it may be necessary to identify a large number of marker loci closely linked to quantitative trait loci (QTL) and then compare genetic distance, based on these linked markers. The use of markers linked to QTLs might result in an association between marker-based estimates of genetic distance and the magnitude of the genetic variance.

Table 5 Pooled genetic variance component estimates, standard errors (SE), and number of environments for yield, plant height, physiological maturity, and lodging for two groups of soybean populations. Group one includes three populations resulting from crossing close-

ly related parents, and group two includes three populations resulting from crossing parents that are assumed unrelated based on coefficient of parentage estimates. Populations were nested within groups

Trait	Group	Genetic variance		Number of environments
		Component	SE	
Yield	One	31 780	5700	4
Yield	Two	58 000	8700	4
Plant height	One	15.63	2.35	4
Plant height	Two	26.12	3.06	4
Physiological maturity	One	10.44	1.10	3
Physiological maturity	Two	13.88	1.47	3
Lodging	One	0.048	0.01	2
Lodging	Two	0.203	0.03	2

The relationship between the genetic variance estimate and the coefficient of parentage does not appear to be of sufficient predictive value for any specific combination of two parents. The average genetic variance of populations derived from closely related parents was smaller than the average genetic variance of populations derived from crosses among PI \times adapted parents.

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